

Review

Role of Vitamin D Metabolism and Activity on Carcinogenesis

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The vitamin D endocrine system regulates a broad variety of independent biological processes, and its deficiency is associated with rickets, bone diseases, diabetes, cardiovascular diseases, and tuberculosis. Cellular and molecular studies have also shown that it is implicated in the suppression of cancer cell invasion, angiogenesis, and metastasis. Sunlight exposure and consequent increased circulating levels of vitamin D are associated with reduced occurrence and a reduced mortality in different histological types of cancer, including those resident in the skin, prostate, breast, colon, ovary, kidney, and bladder. The vitamin D receptor (VDR) as a steroid hormone superfamily of nuclear receptors is highly expressed in epithelial cells at risk for carcinogenesis, providing a direct molecular link by which vitamin D status impacts on carcinogenesis. Because VDR expression is retained in many human tumors, vitamin D status may be an important modulator of cancer progression in persons living with cancer. The aim of this review is to highlight the relationship between vitamin D, VDR, and cancer, summarizing several mechanisms proposed to explain the potential protective effect of vitamin D against the development and progression of cancer.

Key words: Vitamin D; Vitamin D receptor (VDR); Carcinogenesis; Cancer

INTRODUCTION

Laboratory and epidemiological data published over the past several years have contributed to the hypothesis that vitamin D metabolites inhibit cancer development at various tissue sites. In 1937, Peller and Stephenson hypothesized that sunlight exposure reduces the risk of cancer (1), and Apperly demonstrated an association between latitude and cancer mortality in 1941 (2). Four decades later, Garland et al. hypothesized that poor vitamin D status accounts for an elevated risk of colon, breast, and ovarian cancers at higher latitudes in the US (3,4). Schwartz and colleagues hypothesized a similar relationship for prostate cancer (5,6). More recently, Grant demonstrated an inverse correlation between regional type B ultraviolet (UV-B) radiation levels and mortality rates of many cancers, particularly digestive organ cancers, and found that in males approximately 80% of the cancers attributable to low regional solar UV-B were digestive system cancers (7). Mizoue also found an inverse correlation between averaged annual solar radiation levels and mortality from digestive system cancers (i.e., esophagus,

stomach, colon, rectum, pancreas, gallbladder, and bile ducts) but not other cancer types in Japan (8).

THE VITAMIN D SYSTEM

The vitamin D system includes a group of lipid-soluble steroids and their respective metabolites. There are two major forms of vitamin D in nature: ergocalciferol (vitamin D₂) and cholecalciferol (vitamin D₃). Vitamin D₂ is photochemically synthesized in plants or is acquired by a diet of fortified milk products, while vitamin D₃ is produced in the skin of animals and humans in response to sunlight too, in particular to UV-B radiations of appropriate wavelength: 270–300 nm. In most countries in Europe and in the US, the requirement of vitamin D is given by 90% of the 7-dehydrocholesterol cholesterol synthesis in the skin from solar irradiation, and only about 10% is taken up by the diet (9). The classical synthetic pathway involves 25- and 1- α -hydroxylation of vitamin D₂ and D₃ in the liver and kidney, respectively. First, hydroxylation occurs in the liver, and it is led to generate 25(OH)D₃. 25(OH)D₃ enters the systemic circulation, and it has a

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half-life of 12–19 days. Second, hydroxylation occurs in the kidneys, and it constitutes the most biologically active hormonal form of vitamin D: $1,25(\text{OH})_2\text{D}_3$ (calcitriol) (Fig. 1). The serum levels of $25(\text{OH})\text{D}_3$ are a reflection of overall vitamin D status in the body. There are two principal enzymes involved in the formation of circulating

$1,25(\text{OH})_2\text{D}_3$ from dietary absorbed or skin synthesized vitamin D: the hepatic microsomal or mitochondrial vitamin D 25-hydroxylase (CYP27A1) and the renal mitochondrial enzyme 1α -hydroxylase (CYP27B1) for vitamin D and $25(\text{OH})\text{D}_3$, respectively (10). These hydroxylases belong to a class of proteins known as cytochrome P450

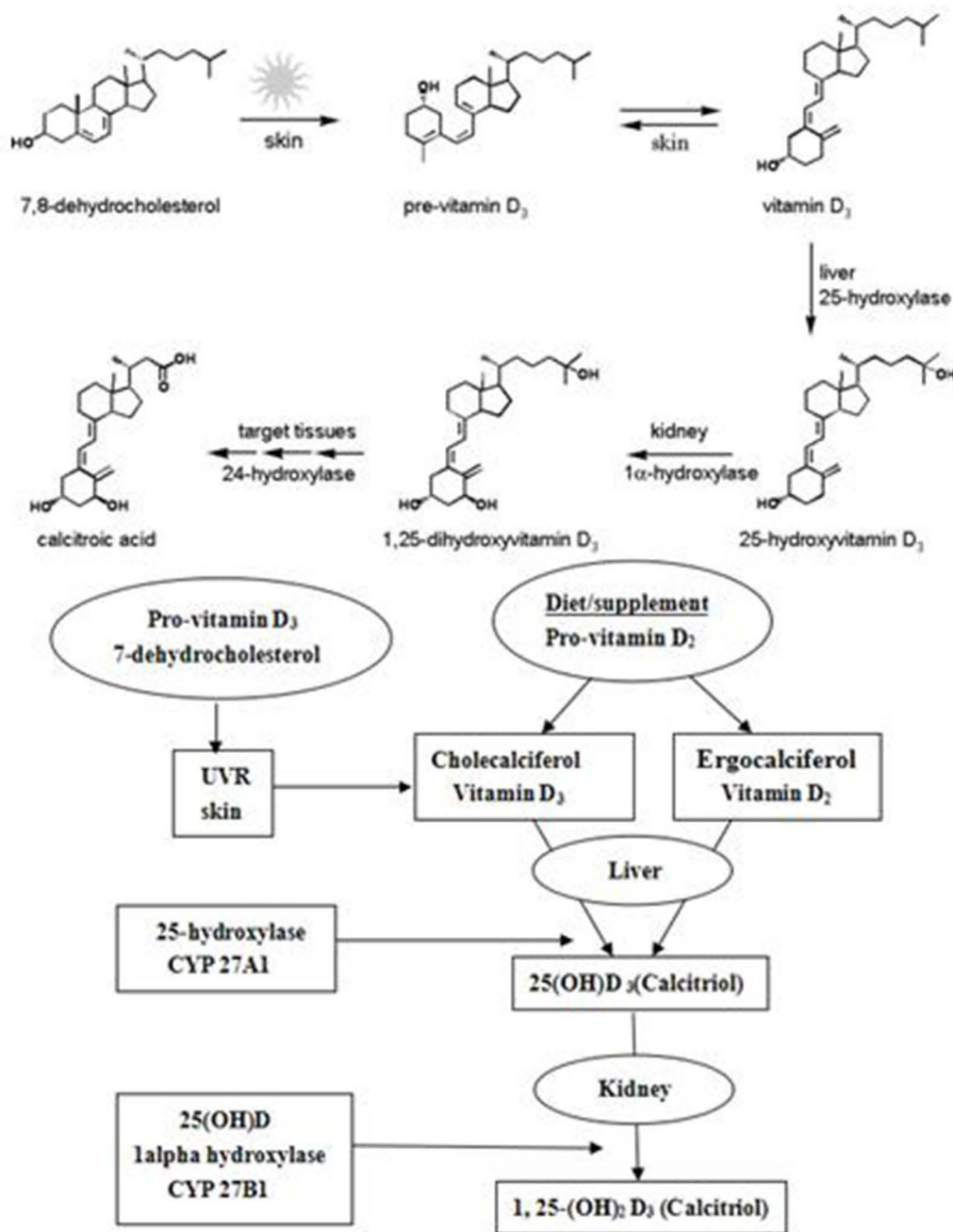


Figure 1. Vitamin D and its metabolites. The vitamin D requirement is from the exposure of skin to sunlight, while a minor portion may be obtained from dietary sources. Upon exposure to ultraviolet B, 7-dehydrocholecalciferol in the skin is photolyzed to form a 9,10-seco-sterol pro-vitamin D₃. Vitamin D₂ (ergocalciferol) or vitamin D₃ (cholecalciferol) made in the skin or ingested in the diet can be stored in and then released from fat cells. The synthetic pathway involves 25- and 1- α -hydroxylation of vitamin D₂ and D₃, in the liver and kidney, respectively. First hydroxylation occurs within the liver and lead to the formation of $25(\text{OH})\text{D}_3$ or calcitriol; second hydroxylation occurs within the kidneys and constitutes the most biologically active hormonal form of vitamin D: $1,25(\text{OH})_2\text{D}_3$ or calcitriol.

mixed function monooxidases. In recent years, extrarenal activity of 25(OH) D_3 -1 α -hydroxylase (CYP27B1) has been reported in various cell types including macrophages, keratinocytes, prostates, and colon cancer cells (11,12). It was shown that 1,25(OH) $_2D_3$ is produced locally in several tissues. It has been demonstrated that potential vitamin D target tissues (e.g., colon, prostate, breast, lung, pancreas) can synthesize and degrade calcitriol. Local production and degradation of calcitriol have been suggested to represent key factors in several types of human cancer (13–15).

THE VITAMIN D RECEPTOR

The vitamin D receptor (VDR) belongs to the superfamily of transacting transcriptional regulatory factors, which includes the steroid and thyroid hormone receptors as well as the retinoid-X receptors and retinoic acid receptors (16). It is an endocrine member of the nuclear receptor superfamily (17) because it is the only nuclear protein that binds the nuclear hormone 1,25(OH) $_2D_3$ with high affinity. The human VDR protein is a 427-amino acid peptide that has a DNA-binding domain, a ligand-binding domain, and activating domains. The VDR protein contains two zinc finger motifs that bind to the DNA, while the ligand-binding domain, located at the carboxy 1 terminus, changes conformation when 1,25(OH) $_2D_3$ binds, allowing interaction with transcription factors. Activated VDR forms a heterodimer with the retinoic acid X receptor, which translocates to the nucleus (18,19) and binds to the vitamin D response element in the promoter region of target genes (20). VDR protein is encoded by a large gene (>100 kb) located on chromosome 12q12-14. The VDR gene encompasses two promoter regions, eight protein-coding exons, and six untranslated exons (21). It has an extensive promoter region capable of generating multiple tissue-specific transcripts. It has been demonstrated that VDR requires heterodimerization with auxiliary proteins for effective DNA interaction.

ROLES OF VITAMIN D AND VITAMIN D RECEPTOR ON CARCINOGENESIS

Several levels of evidence support the relationships among vitamin D, VDR, and cancer: (a) solar UV-B irradiance and vitamin D reduce the risk of incidence and death for many types of cancer, (b) a low intake of vitamin D is associated with an increased risk of cancer; (c) high circulating levels of vitamin D are associated with reduced risk of developing cancer; (d) the aggressiveness of a cancer is lower in summer when the production of vitamin D is higher; (e) polymorphisms of VDR genes affect the risk of developing cancer. These relationships are supported by *in vitro* studies and epidemiologic studies. A lot of *in vitro* studies have demonstrated that exposure of tumor cells to high concentrations of vitamin D compounds inhibits their proliferation and induce differentiation. Numerous

epidemiologic studies have shown the association between factors expected to reduce vitamin D levels (e.g., geography and latitude, history of sun exposure, lifestyle) and the increased rates of cancer, highlighting the protective effects of sunlight and high levels of vitamin D on various types of tumors (2–4,6–9) (Fig. 2).

Colorectal Cancer (CRC)

The ability of 1,25(OH) $_2D_3$ to induce differentiation in colon cancer cells was recognized more than 20 years ago (22), and there is substantial evidence supporting an inverse association between circulating 25(OH) D_3 and CRC risk; meta-analyses and systematic reviews have observed a 50% lower risk of CRC comparing extreme quintiles of 25(OH) D_3 (23,24). Several mechanisms have been hypothesized to underlie this association, some of which may be shared by pathways associated with the putative functional consequences of CRC susceptibility SNPs proximal to VDR DNA binding sites. In addition, vitamin D signaling occurs through binding of the active form 1,25(OH) $_2D_3$ to VDR along specific genomic sequences known as VDREs, which act to activate or repress gene transcription. Several prospective epidemiologic studies, including from this cohort (1), have consistently found an inverse association between higher prediagnostic 25(OH) D_3 levels and CRC risk. Similar to the results for CRC incidence, higher vitamin D levels have been suggested to be inversely associated with CRC-specific and overall mortality among persons diagnosed with CRC in a small number of studies (25–27). Findings from the Nurses' Health Study and the Health Professionals Follow-up Study have shown an association between either higher prediagnostic 25(OH) D_3 levels or higher predicted postdiagnosis 25(OH) D_3 scores and improvement in CRC-specific and overall survival (28). However, one study (29) was limited by its relatively small sample size and the other (30) by its use of predicted, not actual, postdiagnosis vitamin D levels. Another study from Japan has suggested that higher 25(OH) D_3 levels at surgery are associated with a better survival (31), but it is also limited by small sample size.

Breast Cancer

In 1990, Garland et al. first reported an inverse association between total average annual sunlight energy that strikes the ground and age-adjusted breast cancer mortality in the US (4). Several case-control studies have focused on the association between breast cancer risk and circulating levels of 25(OH) D_3 . Results have consistently revealed an inverse association between 25(OH) D_3 and breast cancer (32–34). Other studies have examined the effects of vitamin D on mammary carcinogenesis *in vitro* and in animal models, and the data support a protective role for vitamin D in breast cancer development (35,36). In addition, mice

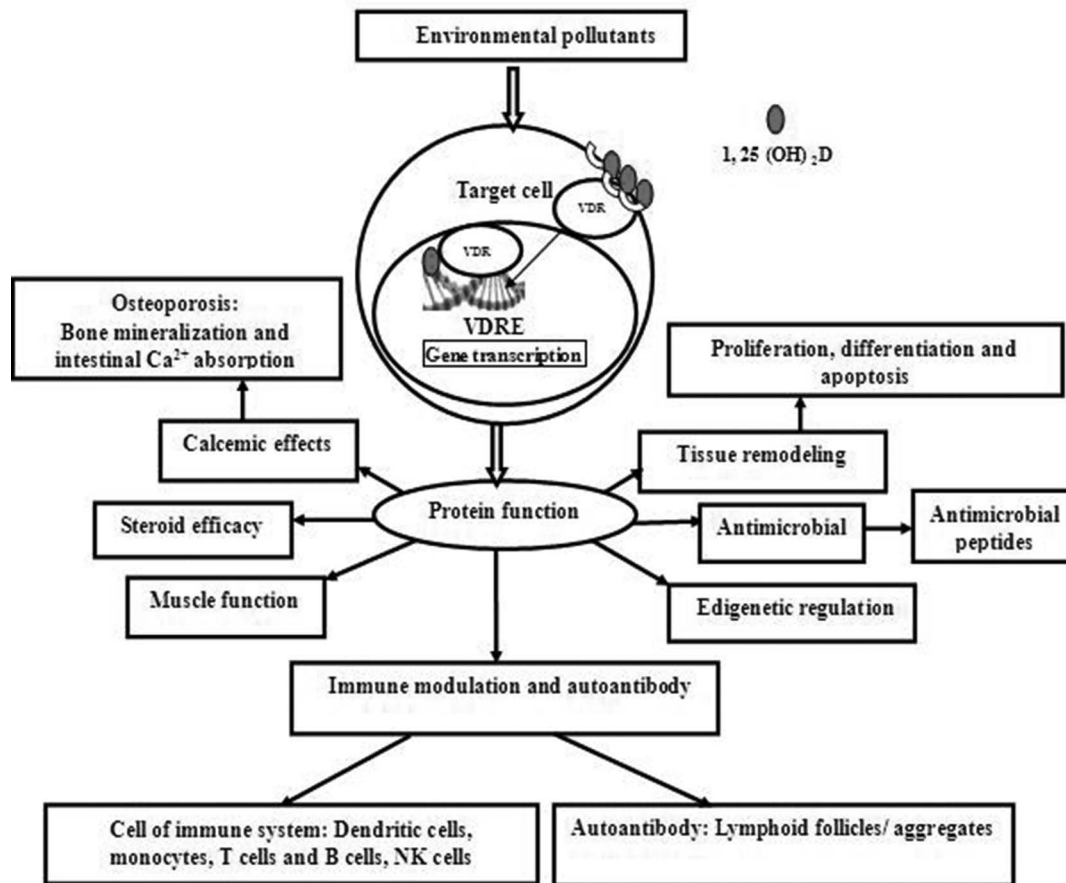


Figure 2. The role of vitamin D/VDR in environmental agent-mediated deregulation. Environmental agents, such as cigarette smoke, particulate matter (less than 10 μm , PM10), ultrafine particles, inhaled oxidants, ozone, and aldehydes activate vitamin D receptor (VDR) and affect different downstream cellular and molecular targets as a result of vitamin D-mediated deregulation. Calcitriol is bound to VDR and vitamin D response elements (VDRE). In conjunction with several transcription factors, this complex led to the transcription of vitamin D-responsive genes. The major cellular and molecular functions affected due to vitamin D/VDR deregulation include calcemic effects, antimicrobial, tissue remodeling, immune modulation and autoantibody production, muscle function, steroid efficacy, epigenetic regulation, immune response, inflammation, and cellular proliferation, differentiation, and apoptosis.

rendered vitamin D deficient exhibit enhanced cancer development (37), as do VDR knockout mice (38). Several mechanisms underlying the inhibitory effects of vitamin D on the growth of breast cancer cells have been proposed. Six case-control studies have examined the relationship between vitamin D intake and breast cancer risk. The largest was an Italian study that included 2,569 cases and 2,588 controls in which a 78-item food frequency questionnaire was used to collect information on dietary sources of vitamin D. Women with the highest vitamin D intake (>190 IU) had a 34% lower risk for breast cancer than those with the lowest vitamin D intake (<60 IU) (39). The odds ratios (ORs) were 0.80 [95% confidence interval (CI) 0.64–0.99] and 0.78 (95% CI 0.66–0.92) among pre- or perimenopausal and postmenopausal women, respectively (40). The strengths of the study are the large dataset and the use of a reproducible and valid food frequency questionnaire

(41). The study results were adjusted for many known risk factors for breast cancer. Limitations of the study include the absence of information on sun exposure or serum levels of vitamin D and the use of hospital-based controls. Two other case-control studies also reported a relatively lower breast cancer incidence with greater vitamin D intake (42). A similar finding was reported in the Women's Health Study cohort that included 10,578 premenopausal women and 20,909 postmenopausal women (43). Higher intake of vitamin D was associated with a lower risk for breast cancer in premenopausal women (OR 0.65; 95% CI 0.42–1.00) but not in postmenopausal women (OR 1.30; 95% CI 0.97–1.13) (44). Other studies that included predominantly postmenopausal women either showed a trend toward a lower breast cancer risk with higher vitamin D intake (45,46) or did not show a protective effect of higher vitamin D intake for breast cancer.

Lung Cancer

In vitro and in vivo studies have demonstrated the antiproliferative effects of $1,25(\text{OH})_2\text{D}_3$ in lung cancer. Higashimoto et al. reported that $1,25(\text{OH})_2\text{D}_3$ inhibited the growth of lung cancer cell lines (47). This effect was mediated by VDR and affected cell cycle regulation in squamous cell carcinoma (SCC) (48). $1,25(\text{OH})_2\text{D}_3$ has also been shown to inhibit lung tumor growth and lung metastases in mouse models (49). Owing to the high number of blood vessels in the lungs, circulating tumor cells easily metastasize there and have proven to be difficult to treat with chemotherapy. Nakagawa et al. demonstrated using Lewis lung carcinoma cells: green fluorescent protein (GFP) construct in a murine model that $1,25(\text{OH})_2\text{D}_3$ strongly inhibited metastatic growth in the lung of VDR null mice (50). In parallel in vitro experiments using Lewis lung carcinoma cells, it was noted that VEGF mRNA, an indicator of angiogenesis, was suppressed following treatment with $1,25(\text{OH})_2\text{D}_3$ at 24 h. The data suggests that $1,25(\text{OH})_2\text{D}_3$ directly reduces tumor metastatic growth in lung cancer cells (51). Several studies reported normal tracheobronchial cells have high levels of 1α -hydroxylase (CYP27B1) enzyme that leads to increased local production of $1,25(\text{OH})_2\text{D}_3$ and low levels of CYP24A1 that leads to increased breakdown. This is in contrast to lung cancer cells that show higher CYP24A1 expression and low to absent CYP27B1. Reciprocal changes that involve an increase in CYP27B1 mRNA and a decrease in CYP24A1 mRNA may play a pivotal role in maintaining the local tissue level of $1,25(\text{OH})_2\text{D}_3$ to be antiproliferative to lung cancer cells (51–53). VDR expression is ubiquitous, and there are data to suggest that higher nuclear VDR expression in lung cancer correlates with improved survival (52). This may relate to increased genomic effects mediated by nuclear VDR on cell cycle-related genes that lead to apoptosis, but this is yet to be confirmed in lung cancer. There are also data to suggest that VDR expression is higher in well-differentiated SCC compared with normal or dysplastic bronchial epithelium (53). This finding is intriguing and worthy of further study to elucidate the relationship between the differentiation status of lung cancer and vitamin D. Chen et al. show a high-level expression of CYP24A1 in subsets of lung cancers and demonstrate an inverse relationship between high CYP24A1 expression and antiproliferative activity of vitamin D (54). Earlier reports regarding increased expression of CYP24A1 in lung adenocarcinoma (55) found that the tumors that had a higher CYP24A1 expression were more poorly differentiated, as well as associated with poor survival. In a parallel in vitro experiment, it was demonstrated that lung cancer cell lines with high CYP24A1 expression had a poorer response to the antiproliferative effects of $1,25(\text{OH})_2\text{D}_3$ compared with those with lower levels of CYP24A1 mRNA. Ramnath et al.

confirmed that CYP24A1 expression was indeed highly expressed in lung cancer compared with nontumorigenic normal bronchial epithelium (56). Analysis of non-small-cell lung carcinoma (NSCLC) cell cultures revealed time-dependent loss of $1,25(\text{OH})_2\text{D}_3$ coincident with the appearance of CYP24A1-generated metabolites. Specific inhibition of CYP24A1 slowed the loss of $1,25(\text{OH})_2\text{D}_3$ and increased the $1,25(\text{OH})_2\text{D}_3$ half-life. These data suggest that increased CYP24A1 expression in lung tumors restricts $1,25(\text{OH})_2\text{D}_3$ antitumor activity.

Prostate Cancer

There is striking geographical variation, such that regional intensity of exposure to solar ultraviolet radiation (UVR) is inversely associated with prostate cancer incidence and mortality in fair-skinned populations (57). Furthermore, inverse associations of cumulative UVR exposure, adult sunbathing, childhood sunburn, and regular holidays in sunny climates with prostate cancer risk have been observed at the individual level (58,59). The effects of UVR on prostate cancer may be mediated by circulating vitamin D levels, the main environmental source of which is sun exposure, which stimulates vitamin D synthesis in the deeper layers of the epidermis. A study based on the Health Professionals Follow-up Study (HPFS) and the Physicians Health Study (PHS) showed that patients with $25(\text{OH})\text{D}_3$ levels <40.5 nmol/L were more likely to die from prostate cancer (HR 1.59, 95% CI 1.06–2.39) compared with levels >95.9 nmol/L (60). From both cohorts, prediagnostic serum samples were used. The association was largely explained by the association between low $25(\text{OH})\text{D}_3$ levels and cancer of advanced stage and higher Gleason score. The association tended to be stronger when restricting the analyses to patients with samples collected within 5 years of the cancer diagnosis. Similar results were observed in a Norwegian study of prostate cancer patients, based on serum samples collected ± 3 months from the date of the cancer diagnosis (61). The risk of cancer death in patients with $25(\text{OH})\text{D}_3$ levels >80 nmol/L was 0.16 (95% CI 0.05–0.43) relative to patients with levels <50 nmol/L. A risk reduction was also seen in patients with $25(\text{OH})\text{D}_3$ levels 50–79 nmol/L (RR 0.33, 95% CI 0.14–0.77). Mice with prostate epithelial cell-specific deletion of VDR (PEC VDRKO) were generated to study the direct effects of VDR on epithelial cell turnover during castration and in response to testosterone repletion. PEC VDRKO mice exhibit lower rates of apoptosis in response to castration and higher rates of proliferation in response to testosterone administration than control mice. These data show that low vitamin D status and VDR deletion alter cell turnover and hormonal responsiveness in normal prostate tissue changes that likely contribute to an increased susceptibility of VDR null mice to PIN and tumorigenesis.

Skin Carcinogenesis

UV induces various types of DNA damage either photochemically or by UV activation of endogenous photoreceptors that create genotoxic free radicals that modify the DNA molecular structure. The most frequently occurring photolesion in sun-exposed human skin is the cyclobutane pyrimidine dimer (CPD) (62,63) particularly thymine dimers, which are induced primarily by UV-B, and also by UV-A to a lesser extent (64,65). CPDs are produced by the dislocation of double bonds in two adjacent pyrimidines by UV absorption, resulting in a cyclobutane ring conformation linking the two nucleobases as a dimer (66,67). Many studies have shown that $1,25(\text{OH})_2\text{D}_3$ reduces thymine dimers in irradiated skin cells in vitro (68) and also in vivo in mouse (69) and human skin (70). Thymine dimers are also reduced in irradiated skin cells in the presence of the low calcemic rapid acting cis-locked nongenomic analogs, $1,25(\text{OH})_2$ -lumisterol₃ (JN) and $1,25(\text{OH})_2$ -7-dehydrocholesterol (JM) in vitro (71) and in mouse skin (69) and also by the transcriptionally active hybrid 1-hydroxymethyl-16-ene-24,24-difluoro-25-hydroxy-26,27-bis-homovitamin D₃. Evidence that the vitamin D photoprotective effect on reductions in thymine dimer DNA damage is via the rapid nongenomic pathway is demonstrated with various vitamin D-like compounds. As noted above, studies by our group have shown that the transcriptionally nonactive $1,25(\text{OH})_2$ -lumisterol₃ protects against UV-induced thymine dimers. Of relevance to the mechanism of action of vitamin D compounds in photoprotection, the coincubation of skin cells with $1,25(\text{OH})_2\text{D}_3$ and 25-dehydro-1 α -hydroxyvitamin D₃-26,23S-lactone (TEI-9647), an antagonist of the genomic action of $1,25(\text{OH})_2\text{D}_3$, did not alter the protective effects of $1,25(\text{OH})_2\text{D}_3$ on thymine dimers. In contrast, coincubation with 1 β , 25-dihydroxyvitamin D₃ (HL), an antagonist of the nongenomic pathway, abolished the photoprotective effect of $1,25(\text{OH})_2\text{D}_3$ (72,73).

Other Tumors

The pathway of vitamin D seems to be involved in the development of endocrine and neuroendocrine tumors too. Studies by Grant as well as by Freedman et al. on cancer mortality rates in the US and Europe, using latitude or DNA-weighted solar UV-B exposure as surrogate endpoints for photoproduction of vitamin D₃ in the skin, found a highly significant association with the incidence of esophagus, stomach, pancreas, bladder, ovary, and uterus, as well as non-Hodgkin lymphoma (3,6,74–75).

THE EFFECT OF VITAMIN D AND CALCIUM ON CARCINOGENESIS

Studies on tissue-specific expression of the CYP27B1-encoded 25-hydroxyvitamin D-1 α -hydroxylase and of the extracellular calcium-sensing receptor (CaR) have led to an understanding of how locally produced $1,25(\text{OH})_2\text{D}_3$

and extracellular calcium act jointly as key regulators of cellular proliferation, differentiation, and function. Thus, impairment of antimitogenic, proapoptotic, and prodifferentiating signaling from the $1,25(\text{OH})_2\text{D}_3$ -activated VDR and from the CaR in vitamin D and calcium insufficiency has been implicated in the pathogenesis of the aforementioned types of cancer. $1,25(\text{OH})_2\text{D}_3$ and calcium interact in modulating cell growth in different ways: (a) signaling pathways from the VDR and the CaR converge on the same downstream elements, for example, of the canonical Wnt pathway; (b) high extracellular calcium modulates extrarenal vitamin D metabolism in favor of higher local steady-state concentrations of $1,25(\text{OH})_2\text{D}_3$; (c) $1,25(\text{OH})_2\text{D}_3$ may upregulate expression of the CaR and thus augment CaR-mediated antiproliferative responses to high extracellular calcium. Grau et al. studied the effect of vitamin D and calcium supplementation on recurrence of colorectal adenomas, who found that calcium supplementation was effective only in patients with normal $25(\text{OH})\text{D}_3$ values (76). Conversely, high $25(\text{OH})\text{D}_3$ levels were associated with a reduced risk of adenoma recurrence only among subjects receiving calcium supplements. Synergistic actions of calcium and vitamin D are probably the reason why high intake of low-fat dairy products is associated with a reduced risk of breast cancer in premenopausal women. Finally, results from studies in animal models of human autoimmune diseases indicated that calcium supplementation was necessary to optimize the therapeutic effect of vitamin D. Therefore, vitamin D, its analogs, and calcium should be further evaluated in clinical trials in patients with early cancer. In the case of established cancer, it is reasonable to consider that combination therapy will be required and that vitamin D, calcium, or an analog added to other effective therapies will likely increase the benefit of the standard therapy and perhaps reduce some of the side effects.

CONCLUSIONS AND FUTURE PERSPECTIVES

This review highlights the relationship between vitamin D, VDR, calcium, and cancer, summarizing several mechanisms proposed to explain the potential protective effect of vitamin D against the development and progression of cancer. It suggests vitamin D, its analogs, and calcium should be further evaluated in clinical trials in patients with early cancer.

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REFERENCES

1. Peller, S.; Stephenson, C. S. Skin irritation and cancer in the United States Navy. *Am. J. Med. Sci.* 194:326–333; 1937.
2. Apperly, F. L. The relation of solar radiation to cancer mortality in North American. *Cancer Res.* 1:191–195; 1941.

3. Garland, C. F.; Garland, F. C. Do sunlight and vitamin D reduce the likelihood of colon cancer? *Int. J. Epidemiol.* 9:227–231; 1980.
4. Garland, F. C.; Garland, C. F.; Gorham, E. D.; Young, J. F. Geographic variation in breast cancer mortality in the United States: A hypothesis involving exposure to solar radiation. *Prev. Med.* 19:614–622; 1990.
5. Schwartz, G. G.; Hulka, B. S. Is vitamin D deficiency a risk factor for prostate cancer? (Hypothesis). *Anticancer Res.* 10:1307–1311; 1990.
6. Hanchette, C. L.; Schwartz, G. G. Geographic patterns of prostate cancer mortality. *Cancer* 70:2861–2869; 1992.
7. Grant, W. B. An estimate of premature cancer mortality in the U.S. due to inadequate doses of solar ultraviolet-B radiation. *Cancer* 94:1867–1875; 2002.
8. Mizoue, T. Ecological study of solar radiation and cancer mortality in Japan. *Health Phys.* 87:532–538; 2004.
9. Norman, A. W. Sunlight, season, skin pigmentation, vitamin D and 25-hydroxy vitamin D: Integral components of the vitamin D endocrine system. *Am. J. Clin. Nutr.* 67:1108–1110; 1998.
10. Cross, H. S. Extrarenal vitamin D hydroxylase expression and activity in normal and malignant cells: Modification of expression by epigenetic mechanisms and dietary substances. *Nutr. Rev.* 65:S108–S112; 2007.
11. Tuohimaa, P. Vitamin D, aging, and cancer. *Nutr. Rev.* 66:S147–S152; 2008.
12. Robsahm, T. E.; Tretli, S.; Dahlback, A.; Moan J. Vitamin D₃ from sunlight may improve the prognosis of breast, colon and prostate cancer (Norway). *Cancer Causes Control* 15:149–158; 2004.
13. Reichrath, J.; Friedrich, M.; Vogt, T. Vitamin D and its analogs in cancer prevention and therapy. *Anticancer Res.* 32:209–210; 2012.
14. Ali, M. M.; Vaidya, V. Vitamin D and cancer. *J. Cancer Res. Ther.* 3:225–230; 2007.
15. Bernardi, R. J.; Johnson, C. S.; Modzelewski, R. A.; Trump, D. L. Antiproliferative effects of 1 alpha, 25-dihydroxy vitamin D(3) and vitamin D analogs on tumor-derived endothelial cells. *Endocrinology* 143:2508–2514; 2002.
16. Chawla, A.; Repa, J. J.; Evans, R. M.; Mangelsdorf, D. J. Nuclear receptors and lipid physiology: Opening the X-files. *Science* 294:1866–1870; 2001.
17. Nuclear Receptor Committee. A unified nomenclature system for the nuclear receptor superfamily. *Cell* 97:161–163; 1999.
18. Polly, P.; Herdick, M.; Moehren, U.; Baniahmad, A.; Heinzl, T.; Carlberg, C. VDR-Alien: A novel, DNA-selective vitamin D₃ receptor-corepressor partnership. *FASEB J.* 14:1455–1463; 2000.
19. Bookout, A. L.; Jeong, Y.; Downes, M.; Yu, R. T.; Evans, R. M.; Mangelsdorf, D. J. Anatomical profiling of nuclear receptor expression reveals a hierarchical transcriptional network. *Cell* 126:789–799; 2006.
20. Rachez, C.; Suldan, Z.; Ward, J.; Chang, C. P.; Burakov, D.; Erdjument, B. H.; Tempst, P.; Freedman, L. P. A novel protein complex that interacts with the vitamin D₃ receptor in a ligand dependent manner and enhances transactivation in a cell-free system. *Genes Dev.* 12:1787–1800; 1998.
21. Dunlop, T. W.; Vaisanen, S.; Frank, C.; Molnar, F.; Sinkkonen, L.; Carlberg, C. The human peroxisome proliferator-activated receptor δ gene is a primary target of 1 α , 25-dihydroxy vitamin D₃ and its nuclear receptor. *J. Mol. Biol.* 349:248–260; 2005.
22. Shirazian, S.; Schanler, M.; Shastry, S.; Dwivedi, S.; Kumar, M.; Rice, K.; Miyawaki, N.; Ghosh, S.; Fishbane, S. The effect of ergocalciferol on uremic pruritus severity: A randomized controlled trial. *J. Ren. Nutr.* 23:308–314; 2013.
23. Yin, L.; Ordonez-Mena, J. M.; Chen, T.; Schottker, B.; Arndt, V.; Brenner, H. Circulating 25-hydroxyvitamin D serum concentration and total cancer incidence and mortality: A systematic review and meta-analysis. *Prev. Med.* 57:753–764; 2013.
24. Autier, P.; Boniol, M.; Pizot, C.; Mullie, P. Vitamin D status and ill health: A systematic review. *Lancet Diab. Endocrinol.* 2:76–89; 2014.
25. Muindi, J. R.; Adjei, A. A.; Wu, Z. R.; Olson, I.; Huang, H.; Groman, A.; Tian, L.; Singh, P. K.; Sucheston, L. E.; Johnson, C. S.; Trump, D. L.; Fakih, M. G. Serum vitamin D metabolites in colorectal cancer patients receiving cholecalciferol supplementation: correlation with polymorphisms in the vitamin D genes. *Horm. Cancer* 4:242–250; 2013.
26. Theodoratou, E.; Palmer, T.; Zgaga, L.; Farrington, S. M.; McKeigue, P.; Din, F. V.; Tenesa, A.; Davey-Smith, G.; Dunlop, M. G.; Campbell, H. Instrumental variable estimation of the causal effect of plasma 25-hydroxy-vitamin D on colorectal cancer risk: Ammendelian randomization analysis. *PLoS One* 7:e37662; 2012.
27. Adams, S. V.; Newcomb, P. A.; Burnett-Hartman, A. N.; White, E.; Mandelson, M. T.; Potter, J. D. Circulating 25-hydroxyvitamin-D and risk of colorectal adenomas and hyperplastic polyps. *Nutr. Cancer* 63:319–326; 2011.
28. Freedman, D. M.; Looker, A. C.; Chang, S. C.; Graubard, B. I. Prospective study of serum vitamin D and cancer mortality in the United States. *J. Natl. Cancer Inst.* 99:1594–1602; 2007.
29. Marian, L. N.; Manson, J. E.; Millen, A.; Pettinger, M.; Margolis, K.; Jacobs, E. T.; Shikany, J. M.; Vitolins, M.; Adams-Campbell, L.; Liu, S.; LeBlanc, E.; Johnson, K. C.; Wactawski-Wende, J. The influence of health and lifestyle characteristics on the relation of serum 25-hydroxyvitamin D with risk of colorectal and breast cancer in postmenopausal women. *Am. J. Epidemiol.* 175:673–684; 2012.
30. Hiraki, L. T.; Qu, C.; Hutter, C. M.; Baron, J. A.; Berndt, S. I.; Bézieau, S.; Brenner, H.; Caan, B. J.; Casey, G.; Chang-Claude, J.; Chanock, S. J.; Conti, D. V.; Duggan, D.; Fuchs, C. S.; Gallinger, S.; Giovannucci, E. L.; Harrison, T. A.; Hayes, R. B.; Hazra, A.; Henderson, B.; Hoffmeister, M.; Hopper, J. L.; Hudson, T. J.; Jenkins, M. A.; Küry, S.; Le Marchand, L.; Lemire, M.; Ma, J.; Manson, J. E.; Nan, H.; Newcomb, P. A.; Ng, K.; Potter, J. D.; Schoen, R. E.; Schumacher, F. R.; Seminara, D.; Slattery, M. L.; Wactawski-Wende, J.; White, E.; Wu, K.; Zanke, B. W.; Kraft, P.; Peters, U.; Chan, A. T. Genetic predictors of circulating 25-hydroxy vitamin D and risk of colorectal cancer. *Cancer Epidemiol. Biomarkers. Prev.* 22:2037–2046; 2013.
31. Otani, T.; Iwasaki, M.; Sasazuki, S.; Inoue, M.; Tsugane, S.; Japan Public Health Center-Based Prospective Study Group. Plasma vitamin D and risk of colorectal cancer: The Japan Public Health Center-Based Prospective Study. *Br. J. Cancer* 97:446–451; 2007.
32. Bauer, S. R.; Hankinson, S. E.; Bertone-Johnson, E. R.; Ding, E. L. Plasma vitamin D levels, menopause, and risk of breast cancer: Dose-response meta-analysis of prospective studies. *Medicine (Baltimore)* 92:123–131; 2013.

33. Slattery, M.; John, E.; Torres-Mejia, G.; Lundgreen, A.; Herrick, J.; Baumgartner, K.; Hines, L.; Stern, M.; Wolff, R. Genetic variation in genes involved in hormones, inflammation, and energetic factors and breast cancer risk in an admixed population. *Carcinogenesis* 33:1512–1521; 2012.
34. Chlebowski, R. T.; Johnson, K. C.; Kooperberg, C.; Pettinger, M.; Wactawski-Wende, J.; Rohan, T.; Rossouw, J.; Lane, D.; O'Sullivan, M. J.; Yasmeen, S.; Hiatt, R. A.; Shikany, J. M.; Vitolins, M.; Khandekar, J.; Hubbell, F. A.; Women's Health Initiative Investigators. Calcium plus vitamin D supplementation and the risk of breast cancer. Women's Health Initiative Investigators. *J. Natl. Cancer Inst.* 100:1581–1591; 2008.
35. Cui, Y.; Rohan, T. E. Vitamin D, calcium, and breast cancer risk: A review. *Cancer Epidemiol. Biomarkers. Prev.* 15:1427–1437; 2006.
36. Jacobs, E. T.; Thomson, C. A.; Flatt, S. W.; Al-Delaimy, W. K.; Hibler, E. A.; Jones, L. A.; Leroy, E. C.; Newman, V. A.; Parker, B. A.; Rock, C. L.; Pierce, J. P. Vitamin D and breast cancer recurrence in the Women's Healthy Eating and Living (WHEL) Study. *Am. J. Clin. Nutr.* 93:108–117; 2011.
37. Tangpricha, V.; Spina, C.; Yao, M.; Chen, T. C.; Wolfe, M. M.; Holick, M. F. Vitamin D deficiency enhances the growth of MC-26 colon cancer xenografts in Balb/c mice. *J. Nutr.* 135:2350–2354; 2005.
38. Miedlich, S. U.; Zhu, E. D.; Sabbagh, Y.; Demay, M. B. The receptor-dependent actions of 1,25-dihydroxyvitamin D are required for normal growth plate maturation in NPT2a knockout mice. *Endocrinology* 151:4607–4612; 2010.
39. Bertone-Johnson, E. R.; Chen, W. Y.; Holick, M. F.; Hollis, B. W.; Colditz, G. A.; Willett, W. C.; Hankinson, S. E. Plasma 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D and risk of breast cancer. *Cancer Epidemiol. Biomarkers Prev.* 14:1991–1997; 2005.
40. Robien, K.; Cutler, G. J.; Lazovich, D. Vitamin D intake and breast cancer risk in postmenopausal women: The Iowa Women's Health Study. *Cancer Causes Control* 18:775–782; 2007.
41. Ross, A. C.; Taylor, C. L.; Yaktine, A. L.; Del valle, H. B. eds. Dietary reference intakes for calcium and vitamin D. Washington, DC: National Academies Press; 2011.
42. Chlebowski, R. T.; Pettinger, M.; Johnson, K. C.; Wallace, R.; Womack, C.; Mossavar-Rahmani, Y.; Stefanick, M.; Wactawski-Wende, J.; Carbone, L.; Lu, B.; Eaton, C.; Walitt, B.; Kooperberg, C. L. Calcium plus vitamin D supplementation and joint symptoms in postmenopausal women in the women's health initiative randomized trial. *J. Acad. Nutr. Diet.* 113:1302–1310; 2013.
43. Patterson, R. E.; Levy, L.; Tinker, L. F.; Kristal, A. R. Evaluation of a simplified vitamin supplement inventory developed for the Women's Health Initiative. *Public Health Nutr.* 2:273–276; 1999.
44. Knight, J. A.; Lesosky, M.; Barnett, H.; Raboud, J. M.; Vieth, R. Vitamin D and reduced risk of breast cancer: A population-based case-control study. *Cancer Epidemiol. Biomarkers. Prev.* 16:422–429; 2007.
45. Millen, A. E.; Wactawski-Wende, J.; Pettinger, M.; Melamed, M. L.; Tylavsky, F. A.; Liu, S.; Robbins, J.; LaCroix, A. Z.; LeBoff, M. S.; Jackson, R. D. Predictors of serum 25-hydroxyvitamin D concentrations among postmenopausal women: The Women's Health Initiative Calcium plus Vitamin D Clinical Trial. *Am. J. Clin. Nutr.* 91:1324–1335; 2010.
46. Anderson, L. N.; Cotterchio, M.; Vieth, R.; Knight, J. A. Vitamin D and calcium intakes and breast cancer risk in pre- and postmenopausal women. *Am. J. Clin. Nutr.* 91:1699–1707; 2010.
47. Higashimoto, Y.; Ohata, M.; Nishio, K.; Iwamoto, Y.; Fujimoto, H.; Uetani, K.; Suruda, T.; Nakamura, Y.; Funasako, M.; Saijo, N. 1 α , 25-dihydroxyvitamin D₃ and all-trans-retinoic acid inhibit the growth of a lung cancer cell line. *Anticancer Res.* 16:2653–2659; 1996.
48. Light, B. W.; Yu, W. D.; McElwain, M. C.; Russell, D. M.; Trump, D. L.; Johnson, C. S. Potentiation of cisplatin anti-tumor activity using a vitamin D analogue in a murine squamous cell carcinoma model system. *Cancer Res.* 57:3759–3764; 1997.
49. Young, M. R.; Ihm, J.; Lozano, Y.; Wright, M. A.; Prechel, M. M. Treating tumor-bearing mice with vitamin D₃ diminishes tumor-induced myelopoiesis and associated immunosuppression, and reduces tumor metastasis and recurrence. *Cancer Immunol Immunother.* 41:37–45; 1995.
50. Nakagawa, K.; Kawaura, A.; Kato, S.; Takeda, E.; Okano, T. 1 α , 25-Dihydroxyvitamin D(3) is a preventive factor in the metastasis of lung cancer. *Carcinogenesis* 26:429–440; 2005.
51. Nakagawa, K.; Sasaki, Y.; Kato, S.; Kubodera, N.; Okano, T. 22-Oxa-1 α ,25-dihydroxyvitamin D₃ inhibits metastasis and angiogenesis in lung cancer. *Carcinogenesis* 26:1044–1054; 2005.
52. Anderson, M. G.; Nakane, M.; Ruan, X.; Kroeger, P. E.; Wu-Wong, J. R. Expression of VDR and CYP24A1 mRNA in human tumors. *Cancer Chemother Pharmacol.* 57:234–240; 2006.
53. Yokomura, K.; Suda, T.; Sasaki, S.; Inui, N.; Chida, K.; Nakamura, H. Increased expression of the 25-hydroxyvitamin D(3)-1 α -hydroxylase gene in alveolar macrophages of patients with lung cancer. *J. Clin. Endocrinol. Metab.* 88:5704–5709; 2003.
54. Chen, G.; Kim, S. H.; King, A. N.; Zhao, L.; Simpson, R. U.; Christensen, P. J.; Wang, Z.; Thomas, D. G.; Giordano, T. J.; Lin, L.; Brenner, D. E. Beer, D. G.; Ramnath, N. CYP24A1 is an independent prognostic marker of survival in patients with lung adenocarcinoma. *Clin. Cancer Res.* 17:817–826; 2011.
55. Jones, G.; Ramshaw, H.; Zhang, A.; Cook, R.; Byford, V.; White, J.; Petkovich, M. Expression and activity of vitamin D-metabolizing cytochrome P450s (CYP1 α and CYP24) in human non small cell lung carcinomas. *Endocrinology* 140:3303–3310; 1999.
56. Ramnath, N.; Nadal, E.; Jeon, C. K.; Sandoval, J.; Colacino, J.; Rozek, L. S.; Christensen, P. J.; Esteller, M.; Beer, D. G.; Kim, S. H. Epigenetic regulation of vitamin d metabolism in human lung adenocarcinoma. *J. Thorac. Oncol.* 9:473–482; 2014.
57. Holmberg, L.; Adolfsson, J.; Mucci, L.; Garmo, H.; Adami, H. O.; Möller, H.; Johansson, J. E.; Stampfer, M. Season of diagnosis and prognosis in breast and prostate cancer. *Cancer Causes Control* 20:663–670; 2009.
58. Schwartz, G. G. Vitamin D and intervention trials in prostate cancer: From theory to therapy. *Ann. Epidemiol.* 19:96–102; 2009.
59. Platz, E. A.; Leitzmann, M. F.; Hollis, B. W.; Willett, W. C.; Giovannucci, E. Plasma 1,25-dihydroxy- and 25-hydroxyvitamin D and subsequent risk of prostate cancer. *Cancer Causes Control* 15:255–265; 2004.
60. Shui, I. M.; Mucci, L. A.; Kraft, P.; Tamimi, R. M.; Lindstrom, S.; Penney, K. L.; Nimsch, K.; Hollis, B. W.;

- Dupre, N.; Platz, E. A.; Stampfer, M. J.; Giovannucci, E. Vitamin D-related genetic variation, plasma vitamin D, and risk of lethal prostate cancer: A prospective nested case-control study. *J. Natl. Cancer Inst.* 104:690–699; 2012.
61. Holt, S. K.; Kwon, E. M.; Koopmeiners, J. S.; Lin, D. W.; Feng, Z.; Ostrander, E. A.; Peters, U.; Stanford, J. L. Vitamin D pathway gene variants and prostate cancer prognosis. *Prostate* 70:1448–1460; 2010.
62. Tatalovich, Z.; Wilson, J. P.; Mack, T.; Yan, Y.; Cockburn, M. The objective assessment of lifetime cumulative ultraviolet exposure for determining melanoma risk. *J. Photochem. Photobiol. B* 85:198–204; 2006.
63. Albarracín, V. H.; Simon, J.; Pathak, G. P.; Valle, L.; Douki, T.; Cadet, J.; Borsarelli, C. D.; Farias, M. E.; Gärtner, W. First characterisation of a CPD-class I photolyase from a UV-resistant extremophile isolated from high-altitude Andean lakes. *Photochem. Photobiol. Sci.* 13:739–750; 2014.
64. Moan, J.; Grigalavicius, M.; Baturaite, Z.; Juzeniene, A.; Dahlback, A. North-South gradients of melanomas and non-melanomas: A role of vitamin D? *Dermatoendocrinology* 5:186–191; 2013.
65. Reichrath, J.; Reichrath, S. The relevance of the vitamin D endocrine system (VDES) for tumorigenesis, prevention, and treatment of non-melanoma skin cancer (NMSC): Present concepts and future perspectives. *Dermatoendocrinology* 5:38–50; 2013.
66. Pescheck, F.; Lohbeck, K. T.; Roleda, M. Y.; Bilger, W. UVB-induced DNA and photosystem II damage in two intertidal green macroalgae: Distinct survival strategies in UV-screening and non-screening Chlorophyta. *J. Photochem. Photobiol. B* 132:85–93; 2014.
67. Emanuele, E.; Spencer, J. M.; Braun, M. From DNA repair to proteome protection: New molecular insights for preventing non-melanoma skin cancers and skin aging. *J. Drugs. Dermatol.* 13:274–281; 2014.
68. Trémezaygues, L.; Seifert, M.; Tilgen, W.; Reichrath, J. 1,25-dihydroxyvitamin D(3) protects human keratinocytes against UV-B-induced damage: In vitro analysis of cell viability/proliferation, DNA-damage and -repair. *Dermatoendocrinology* 1:239–245; 2009.
69. Malley, R. C.; Muller, H. K.; Norval, M.; Woods, G. M. Dietary vitamin D alters the response of the skin to UVB-irradiation depending on the genetic background of the mice. *Photochem. Photobiol. Sci.* 12:536–545; 2013.
70. Gordon-Thomson, C.; Gupta, R.; Tongkao-on, W.; Ryan, A.; Halliday, G. M.; Mason, R. S. 1 α ,25 dihydroxyvitamin D3 enhances cellular defences against UV-induced oxidative and other forms of DNA damage in skin. *Photochem. Photobiol. Sci.* 11:1837–1847; 2012.
71. Wong, G.; Gupta, R.; Dixon, K. M.; Deo, S. S.; Choong, S. M.; Halliday, G. M.; Bishop, J. E.; Ishizuka, S.; Norman, A. W.; Posner, G. H.; Mason, R. S. 1, 25-Dihydroxyvitamin D and three low-calcemic analogs decrease UV-induced DNA damage via the rapid response pathway. *J. Steroid. Biochem. Mol. Biol.* 89:567–570; 2004.
72. Alberg, A. J.; Fischer, A. H. Is a personal history of nonmelanoma skin cancer associated with increased or decreased risk of other cancers? *Cancer Epidemiol. Biomarkers. Prev.* 23:433–436; 2014.
73. Grigalavicius, M.; Juzeniene, A.; Baturaite, Z.; Dahlback, A.; Moan, J. Biologically efficient solar radiation: Vitamin D production and induction of cutaneous malignant melanoma. *Dermatoendocrinology* 5:150–158; 2013.
74. Łuczyska, A.; Kaaks, R.; Rohrmann, S.; Becker, S.; Linseisen, J.; Buijsse, B.; Overvad, K.; Trichopoulos, A.; Valanou, E.; Barmptsoti, A.; Masala, G.; Agnoli, C.; Tumino, R.; Panico, S.; Bueno-de-Mesquita, H. B.; van Duynhoven, F. J.; Peeters, P. H.; Vermeulen, R.; Weiderpass, E.; Brustad, M.; Skeie, G.; González, C. A.; Jakszyn, P.; Quirós, J. R.; Sánchez, M. J.; Huerta, J. M.; Ardanaz, E.; Melin, B.; Johansson, A. S.; Almqvist, M.; Malm, J.; Khaw, K. T.; Wareham, N.; Travis, R. C.; Fedirko, V.; Romieu, I.; Jenab, M.; Gallo, V.; Riboli, E.; Vineis, P.; Nieters, A. Plasma 25-hydroxyvitamin D concentration and lymphoma risk: Results of the European Prospective Investigation into Cancer and Nutrition. *Am. J. Clin. Nutr.* 98:827–838; 2013.
75. Grant, W. B. Sun exposure, vitamin D and cancer risk reduction. *Eur. J. Cancer* 49:2073–2075; 2013.
76. Grau, M. V.; Baron, J. A.; Sandler, R. S.; Haile, R. W.; Beach, M. L.; Church, T. R.; Heber, D. Vitamin D, calcium supplementation, and colorectal adenomas: Results of a randomized trial. *J. Natl. Cancer Inst.* 95:1765–1771; 2003.